

CERTIFICATE OF ANALYSIS

# TaqI

**#ER0673** HC, 15000 u

**Lot:**                      **Expiry Date:**

5'...**T↓C G A**...3'

3'...**A G C↑T**...5'

Concentration: 10 u/μl  
Source: *Thermus aquaticus* YT-1  
Supplied with: 4x1 ml of 10X Buffer TaqI  
1 ml of 10X Buffer Tango™

**Store at -20°C**



In total 6 vials.

BSA included: Lot# BSA62-313P

## RECOMMENDATIONS

**1X Buffer TaqI** (for 100% TaqI digestion)

10 mM Tris-HCl (pH 8.0), 5 mM MgCl<sub>2</sub>, 100 mM NaCl,  
0.1 mg/ml BSA.

**Incubation temperature**

65°C\*.

**Unit Definition**

One unit is defined as the amount of TaqI required to digest 1 μg of lambda DNA *dam*<sup>-</sup> in 1 hour at 65°C in 50 μl of recommended reaction buffer.

**Dilution**

Dilute with Dilution Buffer (#B19): 10 mM Tris-HCl (pH 7.4 at 25°C), 100 mM KCl, 1 mM EDTA, 1 mM DTT, 0.2 mg/ml BSA and 50% glycerol.

**Double Digests**

Tango™ Buffer is provided to simplify buffer selection for double digests. 98% of Fermentas restriction enzymes are active in a 1X or 2X concentration of Tango™ Buffer. Please refer to the Fermentas Catalog or go to [www.fermentas.com/doubledigest](http://www.fermentas.com/doubledigest) to choose the best buffer for your experiments.

1X Tango™ Buffer:

33 mM Tris-acetate (pH 7.9), 10 mM magnesium acetate, 66 mM potassium acetate, 0.1 mg/ml BSA.

\* Incubate under paraffin oil in a capped vial. Incubation at 37°C results in 10% activity.

## Storage Buffer

TaqI is supplied in: 10 mM Tris-HCl (pH 7.5 at 25°C), 300 mM KCl, 1 mM DTT, 0.1 mM EDTA, 0.5 mg/ml BSA and 50% glycerol.

## Recommended Protocol for Digestion

- Add:

nuclease-free water	16 $\mu$ l
10X Buffer TaqI	2 $\mu$ l
DNA (0.5-1 $\mu$ g/ $\mu$ l)	1 $\mu$ l
TaqI	0.5-2 $\mu$ l**
- Mix gently and spin down for a few seconds.
- Incubate under paraffin oil in a capped vial at 65°C for 1-16 hours.

The digestion reaction may be scaled either up or down.

## Recommended Protocol for Digestion of PCR Products Directly after Amplification

- Add:

PCR reaction mixture	10 $\mu$ l (~0.1-0.5 $\mu$ g of DNA)
nuclease-free water	18 $\mu$ l
10X Buffer TaqI	2 $\mu$ l
TaqI	1-2 $\mu$ l**
- Mix gently and spin down for a few seconds.
- Incubate under paraffin oil in a capped vial at 65°C for 1-16 hours.

\*\* This volume of the enzyme is recommended for preparations of standard concentrations (10 u/ $\mu$ l), whereas HC enzymes (50 u/ $\mu$ l) should be diluted with Dilution Buffer to obtain 10 u/ $\mu$ l concentration.

## Thermal Inactivation

TaqI is not inactivated by incubation at 80°C for 20 min.

## Inactivation Procedure

- To prepare the digested DNA for electrophoresis:
  - stop the digestion reaction by adding 0.5 M EDTA, pH 8.0 (#R1021), to achieve a 20mM final concentration. Mix thoroughly, add an electrophoresis loading dye and load onto gel.
- To prepare DNA suitable for further enzymatic reactions:
  - extract with phenol/chloroform, precipitate with ethanol or isopropanol, wash the pellet with 75% cold ethanol and air-dry;
  - dissolve DNA in either nuclease-free water, TE buffer, or a buffer suitable for further applications;
  - check the DNA concentration in the solution.

For **ENZYME PROPERTIES** and **QUALITY CONTROL ASSAY DATA**  
see back page

## ENZYME PROPERTIES

### Enzyme Activity in Fermentas REase Buffers, %

TaqI	B	G	O	R	Tango™	2X Tango™
100	0-20	20-50	20-50	20-50	20-50	20-50

### Methylation Effects on Digestion

Dam: may overlap – blocked.

Dcm: never overlaps – no effect.

CpG: completely overlaps – no effect.

EcoKI: never overlaps – no effect.

EcoBI: never overlaps – no effect.

### Stability during Prolonged Incubation

A minimum of 0.3 units of the enzyme is required for complete digestion of 1 µg of lambda DNA in 16 hours at 65°C.

### Compatible Ends

Bsp119I, Bsu15I, Hin1I, Hin6I, HpaII, MaeII, MspI, NarI, Psp1406I, SsiI, XmiI.

### Number of Recognition Sites in DNA

λ	ΦX174	pBR322	pUC57	pUC18/19	pTZ19R/U	M13mp18/19
121	10	7	4	4	5	12

## QUALITY CONTROL ASSAY DATA

### Overdigestion Assay

No detectable change in the specific fragmentation pattern is observed after a 160-fold overdigestion with TaqI (10 u/µg lambda DNA *dam*<sup>-</sup> x 16 hours).

### Ligation/Recutting Assay

After a 50-fold overdigestion (3 u/µg DNA x 17 hours) with TaqI, more than 95% of the digested DNA fragments can be ligated at a 5'-termini concentration of 1.2 µM. More than 95% of these sites can be recut.

### Labeled Oligonucleotide (LO) Assay

No detectable degradation of single-stranded or double-stranded labeled oligonucleotides occurred during incubation with 10 units of TaqI for 4 hours.

Quality authorized by:

 Jurgita Zilinskiene

### PRODUCT USE LIMITATION.

This product is developed, designed and sold exclusively *for research purposes and in vitro use only*. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals.

Please refer to [www.fermentas.com](http://www.fermentas.com) for Material Safety Data Sheet of the product.